Paul Mooney EMBO Cryo School, Birkbeck College, 9.9.2019

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Getting the most out of a counting direct detector



Talk Plan

- What is counting?
- Getting the most with design.
- Getting the most in operation: what are the choices?
- Getting the most: how do we decide?

- Slides 4-11
- Slides 13-22



Section 1

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What is counting and why do we do it?



All Imaging uses quanta



Siméon Denis Poisson

Before image capture with a camera or an eyeball. So there is something to count if one choses to.



Analog capture of quanta at high dose





Analog capture of quanta at low dose



Electrons as they appear before counting.





Counted capture of quanta at low dose



Electrons counted where they arrived.



Gatan K3 in counting mode



Counted capture of quanta at high dose



Counted image after summing.





Counting reduces image acquisition noise



Analog accumulation



Counted accumulation

"Counting" is done by discrimination

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GALA



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Getting the most out of counting with design



Discriminating electrons from background



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Indirect detectors won't work for counting







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non-CDS

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CDS allows the discrimination threshold to be lowered



Thinning minimizes false positives from scatter



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Direct Detection

High speed minimizes false negatives from coincideence Loss



Li et al, Nature Methods 10, 584–590 (2013).

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Super-resolution by centroiding extracts more location information from each event

1. Electron enters detector



3. Charge collects in each pixel



2. Signal is scattered



4. Events are localized with sub-pixel accuracy



This example shows that centroid locations are plausible



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And these examples show that super-resolution is real and free of artifacts



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K2 Summit in superresolution mode, 1Å super-pixel, 300kV, end of GIF.

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Getting the most out of a counting detector in operation: the options



Choices to make when setting up to image

- Gain correction how long?
- kV
- Dose rate how fast?
- Magnification how fine, how big?



Gain Normalization corrects fixed pattern noise



Same fiberoptic pattern in image and reference





Beware: gain normalization by a noisy gain reference can *create* fixed pattern noise while trying to correct it.



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Uncorrected fixed pattern noise causes self-correlation

Cross-correlated 20e- images with N times that dose in the gain-reference (simulated)





Fixed pattern noise also causes... uh... noise



Without motion correction, high-frequency specimen contrast is destroyed.



With motion correction high-frequency fixed pattern contrast is destroyed because now it is the detector that is drifting.

The stripe of fixed pattern contrast that remains shows that fixed pattern noise IS noise and reduces image quality, even when it isn't obvious.

Images from Li et al, Nature Methods, 2013

Coincidence loss creates mild non-linearity – but the gain reference can still be used over a large range of dose rates



(from Li et al, 2013)

K2 fixed pattern noise fraction after normalization



A 6000 electron gain reference provides < 1% selfcorrelation for > 1 day



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So take a long gain reference at the target dose rate

- If your exposures are to have total dose of 20 electrons, a 2000 electron gain reference will increase noise power by only 1% and yield self-correlations between 20 electron exposures of only 1%
- A 6000 electron gain reference will give extra margin, so that as the gain reference ages over a day or two, the effects will still remain under 1%.
- Dark reference update restores the validity of the gain reference – so the time invested in making the gain reference lasts for much longer than a few days.
- Note that motion correction *does* reduce high-frequency fixed pattern noise, so the gain reference dose requirement may be somewhat lower in practice.

K2 200kV DQE is higher at low spatial frequency



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High resolution being achieved at 200kV

2.6 Å at 200 kV without image filtering or phase plate



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Small-signal AmC SNR implies higher DQE at lower dose rate



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What is the Best Magnification and Binning?



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 300 kV energy-filtered Krios structures from Merk et al, Cell, 2016

- 200 kV Talos Arctica density map from Herzik et al, Nat. Meth., 2017
- *** 300 kV energy-filtered Krios structure, Hong Zhou (private communication)
- **** 200kV, Feathers, Spoth, Fromme, BioArxiv 2019



How do we decide the best parameters?

• DQE – a measure of the camera's efficiency at generating the above kind of SNR from limited dose.

$$DQE(N, s) = \frac{SNR_{out}(N, s)}{SNR_{in}(N, s)}$$

N = incoming average dose in primary electrons / pixel s = spatial frequency

DQE at same (15 e/pix/s) dose rate for non-CDS and CDS



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Nyquist fraction	absolute increase	relative increase
03	0-5%	0-6%
1	7%	28%
1.25	5%	38%

DQE allows specimen- and microscope- independent camera characterization as for the K3 shown here



Higher magnification maps specimen contrast to lower spatial frequencies on the camera – where the DQE is higher



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17may06a_b_00012gr_00020sq_v01_00004h116_00006edhii-a-Dk

100mm

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But this may not be an issue for small tightlypacking molecules

Aldolase image courtesy of Gabriel Lander and Mark Herzik



Tradeoff Summary: Image Quality vs Throughput

- Lower kV improves beam-specimen interaction but affects spatial frequencies at the camera differently, raising low frequency DQE and lowering high frequency DQE
- Higher dose rate reduces exposure time but also reduces image quality as measured by DQE.
- Higher magnification increases the DQE with which specimen contrast is detected but at the expense of field of view.

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Getting the most out of a counting detector in operation: how to decide



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structures / day

Image quality is also "throughput"



Rosenthal and Henderson, JMB, 2003, figure 11.



Rosenthal-Henderson plot of Feather et al table 2



Higher mag has lower B-factor – but is benefit outweighed by slower particle accumulation?

Derived from Feathers et al, BioArxiv, 6/19/19, table 2

Rosenthal-Henderson plot redrawn vs image count



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Higher mag converges faster even with fewer particles per frame.

Derived from Feathers et al, BioArxiv, 6/19/19, table 2 using table 1 average particles per image



→ Use B-factor for optimization of real throughput

3.0



- Puts image quality and "molecules per day" into the correct relationship.
- Can include system issues like grid materials, algorithm varients, etc.
- Small particle counts could tell which line one's parameter choices (A, B or C) lie on without a full-out 1M particle data set

2.2

Reading List

- Chiu, et al, Evaluation of super-resolution performance of the K2 electron-counting camera using 2D crystals of aquaporin-0, JSB 2015.
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Backup

3.8Å reconstruction from one K3 image



Surpassing the physical Nyquist limit to produce super-resolution cryo-EM reconstructions

J. Ryan Feathers¹, Katherine A. Spoth², and J. Christopher Fromme¹

bioRxiv preprint first posted online Jun. 19, 2019; doi: http://dx.doi.org/10.1101/675397.

super

Nyquist

limit